



C3H-10T1-2

CSI370Mu11

Instruction manual

FOR RESEARCH USE ONLY
NOT FOR USE IN CLINICAL DIAGNOSTIC PROCEDURES

1st Edition (Revised in Nov, 2023)

[DESCRIPTION]

C3H-10T1-2 is a cell line exhibiting fibroblast morphology that was isolated from a line of C3H mouse embryo cells. This cell line was deposited by C Heidelberger.

Synonyms: C3H/10T1/2; C3H/10T1/2-clone8; C3H/10T1/2 CL8; C3H10T1-2; C3H10T1/2; C3H-10T1/2;

Organism: C3H mouse

Tissue: Embryo

Disease: Sarcoma

Cell Type: Fibroblast

Growth properties: Adherent

[PROPERTIES]

Cell activity: >95% (Viability by Trypan Blue Exclusion).

Formulation: Frozen 1 mL or T25 flask.

Biosafety: Negative for HIV-1, HBV, HCV, mycoplasma, bacteria, yeast and fungi.

Applications: For research use only. It is not approved for human or animal use, or for application in clinical diagnostic procedures.

Size: >5×10⁵cell/vial

[STORAGE]

Upon receiving, directly and immediately transfer the cells from dry ice to liquid nitrogen and keep the cells in liquid nitrogen until they are needed for experiments.

Form & Buffer: Supplied as solution form in frozen stock solution, containing 50% base medium +40%FBS+10%DMSO.

Storage conditions: liquid nitrogen

[USAGE]

Culture conditions:

Complete growth medium: DMEM+10%FBS+1%Penicillin-Streptomycin Solution

Temperature: 37°C

Condition: 95% air, 5% carbon dioxide



Cell recovery:

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the cap out of the water. The thawing time is about 2 minutes.
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by spraying with 75% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 9.0mL complete culture medium. and spin at approximately 1000 rpm for 5 minutes.
4. Resuspend cell pellet with the recommended complete medium . and dispense into a T25 culture flask.
5. Incubate the culture at 37°C, 5% CO₂ in a suitable incubator.

Cell passage:

1. Cell passage when cell growth at 85-95%.
2. Remove and discard culture medium and wash with PBS 1-2 times.
3. Add 1.0 mL of Trypsin-EDTA solution to flask and observe cells under an inverted microscope until cell layer is dispersed (usually within 2 to 3 minutes. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal). Stop digestion by adding 2-3 ml of complete medium containing 10% serum. Make it a single cell suspension.
4. Add the fresh medium to resuspend the cells. Unless otherwise stated, the recommended ratio of primary cells is 1/3-1/4.

[Shipping]

Dry ice.

[IMPORTANTNOTE]

1. The cell is for research use only, and we will not be responsible for any issue if the cell was used in clinical diagnostic or any other procedures.
2. Read the instructions carefully, and keep and operate in strict accordance with the instructions.
3. After cell recovery, please take regular microscopic examination and photos to record the growth status of cells.
4. If you observe abnormalities or have questions about cell culture operations, please contact us in time.